Clinically Relevant Plasma Concentrations of Colistin in Combination with Imipenem Enhance Pharmacodynamic Activity against Multidrug-Resistant *Pseudomonas aeruginosa* at Multiple Inocula

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The use of combination antibiotic therapy may be beneficial against rapidly emerging resistance in *Pseudomonas aeruginosa*. The aim of this study was to systematically investigate *in vitro* bacterial killing and resistance emergence with colistin alone and in combination with imipenem against multidrug-resistant (MDR) *P. aeruginosa*. Time-kill studies were conducted over 48 h using 5 clinical isolates and ATCC 27853 at two inocula (~10^6 and ~10^8 CFU/ml); MDR, non-MDR, and colistin-heteroresistant and -resistant strains were included. Nine colistin-imipenem combinations were investigated. Microbiological response was examined by log changes at 6, 24, and 48 h. Colistin combined with imipenem at clinically relevant concentrations increased the levels of killing of MDR and colistin-heteroresistant isolates at both inocula. Substantial improvements in activity with combinations were observed across 48 h with all colistin concentrations at the low inoculum and with colistin at 4× and 16× MIC (or 4 and 32 mg/liter) at the high inoculum. Combinations were additive or synergistic against imipenem-resistant isolates (MICs, 16 and 32 mg/liter) at the 10^6-CFU inoculum in 9, 11, and 12 of 18 cases (i.e., 9 combinations across 2 isolates) at 6, 24, and 48 h, respectively, and against the same isolates at the 10^8-CFU inoculum in 11, 7, and 8 cases, respectively. Against a colistin-resistant strain (MIC, 128 mg/liter), combinations were additive or synergistic in 9 and 8 of 9 cases at 24 h at the 10^6- and 10^8-CFU inocula, respectively, and in 5 and 7 cases at 48 h. This systematic study provides important information for optimization of colistin-imipenem combinations targeting both colistin-susceptible and colistin-resistant subpopulations.

The world is facing a growing threat from multidrug-resistant (MDR) Gram-negative “superbugs,” such as *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, and *Klebsiella pneumoniae* (19, 30, 50). This problem has increased substantially over the past 5 years, especially for infections by these pathogens (1, 27, 35), and therefore, its use has increased substantially over the past 5 years, especially for critically ill patients (6, 27).

It is now evident that the concentrations of colistin achieved in the plasma of critically ill patients with the currently recommended dosage regimens are suboptimal for a significant proportion of patients (13, 43). Unfortunately, increasing the daily dose may not be an acceptable option, since nephrotoxicity is a dose-limiting adverse effect and occurs in 30 to 50% of patients (13, 16, 22). It is therefore not surprising that suboptimal concentrations cause the emergence of resistance to colistin, which seriously threatens colistin therapy (45, 48). *In vivo* (21, 33) and *in vitro* (3, 4) studies show the potential for the rapid emergence of colistin resistance with monotherapy. The phenomenon of colistin heteroresistance (the presence of colistin-resistant subpopulations in an isolate considered susceptible by MIC measurement) (56) has been reported for *A. baumannii* (29, 56) and *K. pneumoniae* (34, 44, 54) but not yet for *P. aeruginosa*. Heteroresistance very likely contributes to the emergence of colistin resistance. The aim of the present study was to systematically investigate the extent of *in vitro* bacterial killing and the emergence of colistin resistance with colistin alone and in combination with imipenem against *P.


**TABLE 1. MICs for the *P. aeruginosa* isolates used in this study**

<table>
<thead>
<tr>
<th>Isolate</th>
<th>MIC (mg/liter)*</th>
<th>Cephalosporinase and carbapenemase typing</th>
<th>MDRb</th>
</tr>
</thead>
<tbody>
<tr>
<td>ATCC 27853</td>
<td>2</td>
<td>Negative</td>
<td>No</td>
</tr>
<tr>
<td>19147 n/m</td>
<td>128</td>
<td>IMP and CTX-M positivec</td>
<td>Yes</td>
</tr>
<tr>
<td>20509 n/m</td>
<td>0.5</td>
<td>Negative</td>
<td>Yes</td>
</tr>
<tr>
<td>19271 n/m</td>
<td>2</td>
<td>Negative</td>
<td>Yes</td>
</tr>
<tr>
<td>20891 n/m</td>
<td>1</td>
<td>Negative</td>
<td>Yes</td>
</tr>
</tbody>
</table>

*a* CLSI breakpoints for colistin were ≤2 mg/liter for susceptibility, 4 mg/liter for intermediacy, and ≥8 mg/liter for resistance. For imipenem, the breakpoints were ≤4 mg/liter for susceptibility, 8 mg/liter for intermediacy, and ≥16 mg/liter for resistance (10).

*b* Defined as diminished susceptibility to ≥2 of the following 5 drug classes: antipseudomonal cephalosporins, antipseudomonal carbapenems, β-lactam-β-lactamase inhibitor combinations, antipseudomonal fluoroquinolones, and aminoglycosides (40).

*c* Colistin heteroresistant. Heteroresistance to colistin was defined as the existence, in an isolate for which the colistin MIC was ≤2 mg/liter, of subpopulations able to grow in the presence of >2 mg/liter colistin (55). Contains genes encoding an IMP-type carbapenemase and a CTX-M-type ESBL.

### MATERIALS AND METHODS

#### Bacterial isolates

Five clinical isolates and *P. aeruginosa* ATCC 27853 (American Type Culture Collection, Manassas, VA) were selected to represent a mixture of strains susceptible and resistant to colistin and imipenem, colistin-heteroresistant and nonheteroresistant strains, and multidrug-resistant (MDR) and non-MDR strains. MDR was defined as diminished susceptibility to at least two of the following five drug classes: antipseudomonal cephalosporins, antipseudomonal carbapenems, β-lactam-β-lactamase inhibitor combinations, antipseudomonal fluoroquinolones, and aminoglycosides (40). In addition, all strains were examined by PCR for the presence of genes encoding cephalosporinases and carbapenemases, i.e., IMP-, VIM-, NDM-, KPC-, CTX-M-, SHV-, and CMY-type β-lactamases (47, 58). The isolates are described in detail in Table 1. All clinical isolates were collected from patients with cystic fibrosis, had different susceptibilities to colistin and imipenem, and combination to 20 ml of a log-phase broth culture of approximately 106 or 108 (primarily 108) CFU/ml. For monotherapy with colistin or imipenem, 2-fold multiples of the MIC (0.25× to 64× MIC) were employed for susceptible isolates. For the colistin-resistant isolate (19147 n/m; MIC, 128 mg/liter), a single colistin concentration of 32 mg/liter was employed. Imipenem concentrations of 1, 8, and 32 mg/liter were used for imipenem-resistant isolates. In combination experiments, both antibiotics were studied at concentrations of 0.5×, 4×, and 16× MIC for susceptible isolates; resistant isolates, concentrations of 1, 4, and 32 mg/liter for colistin and 1, 8, and 32 mg/liter for imipenem were employed. In total, nine colistin-imipenem combinations were examined for each isolate at each inoculum. Prior to each experiment, isolates were subcultured onto horse blood agar (Media Preparation Unit) and were incubated at 35°C overnight. One colony was then selected and grown overnight in 10 ml CAMHB at 37°C; from this colony, an early-log-phase culture was obtained. Each antibiotic was added alone or in combination to 20 ml of a log-phase broth culture of approximately 106 or 108 CFU/ml to yield the desired concentrations. Each 20-ml culture was placed in a sterile 50-ml polypropylene tube (Greiner Bio One) and was incubated in a shaking water bath at 37°C. Serial samples (100 µl) were collected aseptically for viable-cell counting at 0, 0.5, 1, 2, 4, 6, 8, and 24 h and for PAPs at 48 h (see above) for all experiments involving colistin (including combination arms) and for viable-cell counting at 0, 1, 2, 4, 6, 24, and 48 h for experiments with imipenem alone. Immediately after sampling and serial dilution, 50 µl of the bacterial cell suspension was spirally plated onto nutrient agar with enumeration after 24 h of incubation (48 h for plates with small colonies) as described under “PAPs” above.

**Pharmacodynamic (PD) analysis.** Microbiological responses to monotherapy and combination therapy were examined using the log change method, comparing the change in bacterial counts (log10 CFU/ml) from that at 0 h [log10(CFU0)] to that at a given time (t) [log10(CFUt)] to log10(CFU0) − log10(CFUt) as follows: log change = log10(CFU0) − log10(CFUt).

Single antibiotic or combination regimens causing a reduction of ≥1 log10 CFU/ml from the initial inoculum at 6, 24, or 48 h were considered active. We considered synergy to be indicated by a ≥2-log10 CFU/ml-lower bacterial count with the combination than with its most active component at the specified time (42); additivity was defined by a 1- to <2-log10 CFU/ml-lower bacterial count with the combination.

#### RESULTS

**Microbiological response.** The various susceptibilities of the isolates to colistin are evident in the PAPs obtained prior to...
achieving a ~2-log_{10} kill over that with active monotherapy at 6 h. Against all three isolates, there were 10 and 9 cases of additivity/synergy at 24 and 48 h, respectively, mostly involving colistin at 4× or 16× MIC (Table 2).

**Imipenem-resistant isolates.** For the two imipenem-resistant isolates (19271 n/m and 20891 n/m), there was no evidence of carbapenemase activity; most likely, an alternative resistance mechanism, such as the loss of major outer membrane proteins, was present. At the low inoculum, combination therapy resulted in substantial improvements in bacterial kill with all colistin concentrations across 48 h. At 6 h, additivity/synergy occurred in 9 of 18 cases (i.e., 9 combinations across 2 isolates), predominantly against isolate 19271 n/m; additivity/synergy occurred with combinations containing colistin at all concentrations and produced additional reductions of ~2- to 6-log_{10} CFU/ml over that with usually active colistin monotherapy (Table 3 and Fig. 2). In 5 of 6 cases involving colistin at 4× or 16× MIC against 19271 n/m, bacterial counts were reduced to below the limit of detection (i.e., 20 CFU/ml). Substantial improvements in activity against both isolates were also observed at 24 and 48 h at all colistin concentrations. Additivity/synergy occurred in 11 and 12 of 18 cases at 24 and 48 h, respectively, resulting in an additional Δ1- to 4-log_{10} kill at 24 h and >2.5-log_{10} kill at 48 h over that with monotherapy (Table 3 and Fig. 2). Interestingly, the combinations of colistin at 0.5×, 4×, or 16× MIC with imipenem at 32 mg/liter each reduced the bacterial loads of both isolates to below the limit of detection at 24 h; the maximum reduction in bacterial counts (log_{10} CFU/ml) at 24 h with colistin monotherapy at 16× MIC was ~4.5. Improvements in activity with combination therapy at the high inoculum also occurred at all time points but were essentially restricted to combinations containing colistin at 4× or 16× MIC. Ten of 12 cases at 6 h containing colistin at 4× MIC (Table 3) or 16× MIC (data not shown) showed additivity or synergy. At 24 and 48 h, the addition of imipenem at all concentrations to colistin at 4× or 16× MIC produced additivity/synergy in more than half of all cases and substantially improved the activity over that with each antibiotic alone (by as much as Δ4-log_{10} kill).

**Colistin-resistant isolate.** Bacterial killing at the 10^{6}-CFU/ml inoculum was substantially enhanced at 24 h: all combinations tested were additive or synergistic, and only one combination (colistin at 1 mg/liter plus imipenem at 0.5× MIC) was inactive (Table 3). The addition of all colistin concentrations to imipenem at 4× or 16× MIC produced Δ3.5- to 4.5-log_{10} kill at 24 h, substantially higher than that with equivalent imipenem monotherapy. At 48 h, all colistin concentrations in combination with imipenem at 4× MIC were synergistic (~2- to 4-log_{10} kill) and substantially improved activity over that with equivalent monotherapy. The addition of colistin at 32 mg/liter to imipenem (all concentrations) was additive or synergistic at a substantially earlier time (6 h), with Δ1- to 2-log_{10} greater kill than that with equivalent imipenem monotherapy (overall kill, Δ3- to 5-log_{10} CFU/ml). At the high inoculum, additivity was achieved at 6 h with all combinations containing colistin at 4 mg/liter (Table 3) and 32 mg/liter (data not shown), and activity was enhanced by Δ1-log_{10} kill over that with imipenem monotherapy. Eight of 9 combinations at 24 h and 7 of 9 combinations at 48 h were additive or synergistic, encompassing all colistin concentrations and in many cases

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**FIG. 1.** Baseline PAPs of the reference strain and all clinical isolates at an initial inoculum of ~10^{5} CFU/ml. The y axis starts from the limit of detection, and the LOQ is indicated by the dashed horizontal line.

colistin treatment (Fig. 1). Representative time-kill profiles for colistin and imipenem monotherapy and combination therapy are shown in Fig. 2 (inoculum, ~10^{5} CFU/ml) and 3 (inoculum, ~10^{6} CFU/ml). Log changes in viable cell counts at each inoculum with clinically relevant colistin concentrations are presented in Tables 2 and 3. Additional time-kill and log change data are presented in the supplemental material. At the 10^{6}-CFU/ml inoculum, regrowth was observed to various extents for all susceptible isolates at 48 h with colistin monotherapy with the majority of concentrations. Regrowth with imipenem monotherapy at concentrations of ≥4× or 8× MIC was more variable and substantially less for susceptible isolates at 48 h, even when extended-spectrum β-lactamases (ESBLs) were present. An inoculum effect with colistin monotherapy was generally observed (Fig. 2 and 3, left). Killing by imipenem was generally slightly slower at the high inoculum than at the low inoculum, although the extents of reduction in bacterial counts (log_{10} CFU/ml) were comparable at the two inocula (Fig. 2 and 3).

**Isolates susceptible to both colistin and imipenem.** At the 10^{6}-CFU/ml inoculum, the addition of colistin at 0.5× MIC to imipenem (all concentrations) resulted in additivity or synergy at 6 h in 7 of 9 cases (i.e., 3 combinations against 3 isolates), achieving a ~2- to 3-log_{10} greater kill than that with the most active equivalent monotherapy, and undetectable bacterial counts in many cases (Table 2 and Fig. 2). By 24 or 48 h, improvements in activity with combination therapy over that with the most active monotherapy (usually imipenem) were modest, particularly when only clinically relevant concentrations of colistin (0.5× or 4× MIC) were considered. Of the 27 cases (i.e., 9 combinations against 3 isolates), 7 at 24 h and 8 at 48 h showed additivity or synergy, although only 1 case resulted in activity (i.e., ≥1-log_{10} kill) if equivalent monotherapy with either drug was inactive. A similar pattern of activity was observed at the 10^{5}-CFU/ml inoculum. For ATCC 27853, combinations containing colistin at 4× MIC provided an additional

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resulting in additional reductions of $-1 \log_{10}$ to $4 \log_{10}$ CFU/ml over that with the most active monotherapy (imipenem at 16× MIC). This enhancement of activity was particularly evident with combinations containing colistin at 4 or 32 mg/liter, and on two occasions, when colistin was combined with imipenem at 16× MIC, no viable bacteria were detected at 48 h.

**Emergence of colistin resistance.** For the 4 colistin-heteroresistant isolates (Table 1), the proportion of resistant subpopulations at 10^8 CFU/ml ranged from $2.2 \times 10^{-7}$ to $4.7 \times 10^{-3}$ (Fig. 1). With colistin monotherapy against isolates susceptible to both colistin and imipenem, real-time PAPs performed at 48 h in the time-kill studies demonstrated increases in colistin-resistant subpopulations at both the low and high inocula with clinically relevant colistin concentrations (examples are shown in Fig. 2 and 3; see also the supplemental material); no such increase was observed with isolate 19056 muc at the high inoculum. Against imipenem-resistant isolate 19271 n/m, colistin concentrations of 0.25 to 64× MIC at the low inoculum and 1× to 64× MIC at the high inoculum resulted in nearly 100% of the remaining cells at 48 h growing in the presence of 10 mg/liter colistin. In contrast, no increase in colistin-resistant subpopulations was observed for the imipenem-resistant isolate 20891 n/m at either inoculum. Combi-
nation therapy against colistin-susceptible isolates generally had little effect on the proportion of colistin-resistant subpopulations at 48 h at either inoculum; the shapes of the PAPs were very similar to those obtained with equivalent colistin monotherapy (Fig. 2 and 3).

DISCUSSION

Although colistin has been commercially available for more than 50 years (27), reliable PK/PD data have emerged only recently. Population PK studies have shown that plasma colistin concentrations achieved with currently recommended CMS dosage regimens are likely to be suboptimal for many patients, typically generating average steady-state plasma colistin concentrations of ~2 to 3 mg/liter, with some patients achieving concentrations as high as ~10 mg/liter (13, 18, 24, 28, 32, 43). Increasing the daily dose of CMS for such patients may not be an option, since nephrotoxicity, which occurs in ~30 to 50% of patients (16, 22), is a dose-limiting adverse effect. Given these circumstances and the current last-line status of colistin therapy, we chose to examine not only synergy but also additivity, since even a relatively small increase in activity with combination therapy may be beneficial for patient care. Because colis-
tin is almost entirely unbound in CAMHB (3), colistin concentrations of 0.5 \( \mu \)g/liter and 4 \( \mu \)g/liter for isolates with MICs of 1 \( \mu \)g/liter and 16 \( \mu \)g/liter (1 and 4 \( \mu \)g/liter for colistin-resistant isolates) used in our study are clinically relevant, even assuming that binding of colistin by plasma in patients is similar to that in animals (i.e., 50% bound) (25). Considering the effect of protein binding, all the imipenem concentrations employed are readily achieved in plasma (49).

Because some data show that the activities of both colistin (8) and imipenem (36) are attenuated at high inocula compared to those at low inocula, experiments were conducted at inocula of both \( 10^6 \) and \( 10^8 \) CFU/ml. An inoculum effect was generally observed for colistin monotherapy, whereas no obvious inoculum effect was present for imipenem (Fig. 2 and 3). Regrowth of all isolates was observed with colistin monotherapy, even at colistin concentrations well above those that can safely be achieved clinically. Similar regrowth with colistin (or polymyxin B) monotherapy has been observed for colistin-resistant \( P. \) aeruginosa both in vitro (4, 8, 15, 51) and in vivo (21). For \( A. \) baumannii and \( K. \) pneumoniae, regrowth following colistin monotherapy has been attributed to the amplification of colistin-resistant subpopulations (11, 44, 52), with colistin heteroresistance reported in both species (17, 29, 44, 54). We have reported here, for the first time, colistin heteroresistance in \( P. \) aeruginosa.

The emergence of colistin resistance following colistin monotherapy has been reported previously for \( P. \) aeruginosa at both low and high inocula (4, 8), and a similar phenomenon was observed in the present study for all isolates except 20981 n/m. While \( P. \) aeruginosa can undergo adaptive resistance to polymyxins (14), the presence of colistin heteroresistance at baseline and the changes in PAPs after treatment suggest that regrowth following colistin monotherapy may be due to amplification of pre-existing colistin-resistant subpopulations. This possibility suggests that care is required with colistin monotherapy against \( P. \) aeruginosa, even where isolates appear susceptible on the basis of MICs.

The addition of imipenem to colistin at both inocula generally resulted in substantial improvements in bacterial killing over that with equivalent monotherapy against MDR \( P. \) aeruginosa isolates resistant to either antibiotic, even when ESBLs were present. The improvements in activity against these isolates were observed across the 48-h duration, with all colistin concentrations at the low inoculum and with colistin at 4 \( \mu \)g/liter and 16 \( \mu \)g/liter (MIC for both antibiotics) at the high inoculum. Notably, the total reductions in bacterial counts (log_{10} CFU/ml) achieved with combinations containing lower colistin concentrations (0.5 \( \mu \)g/liter and 4 \( \mu \)g/liter or 1 and 4 \( \mu \)g/liter) were on many occasions similar in magnitude to the reductions achieved with combinations containing colistin at 16 \( \mu \)g/liter, particularly at the \( 10^6 \)-CFU/ml inoculum (Table 3). This suggests that combinations of colistin and imipenem containing clinically relevant colistin concentrations may be as effective as combinations containing higher concentrations against MDR isolates when resistance to either drug is present. This is an important result, given that colistin-induced nephrotoxicity is a dose-limiting adverse effect.

The benefits of the addition of imipenem to colistin for overall antibacterial activity were less pronounced against the three isolates susceptible to both antibiotics and were generally restricted to improvements in initial kill, i.e., up to 6 h (Table 2). Because a proportion of patients will achieve only low plasma colistin concentrations with the currently recommended dosage regimens (13, 43), the combination of colistin

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**Table 2.** Log changes in viable cell counts at 6, 24, and 48 h with various clinically relevant concentrations of colistin and imipenem against three \( P. \) aeruginosa isolates susceptible to both antibiotics

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Inoculum (CFU/ml)</th>
<th>Time (h)</th>
<th>Col 0.5 × MIC</th>
<th>Col 4 × MIC</th>
<th>Imi 0.5 × MIC</th>
<th>Imi 4 × MIC</th>
<th>Log change [log_{10}(CFU/ml) - log_{10}(CFU/ml)]</th>
</tr>
</thead>
<tbody>
<tr>
<td>ATCC</td>
<td>( 10^6 )</td>
<td>6</td>
<td>-0.41</td>
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</tr>
<tr>
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</tr>
<tr>
<td></td>
<td></td>
<td>48</td>
<td>3.80</td>
<td>+1.35</td>
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</tr>
<tr>
<td>27853</td>
<td>( 10^8 )</td>
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<td>-0.06</td>
<td>-2.04</td>
<td>2.18</td>
</tr>
<tr>
<td></td>
<td></td>
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<td>+1.67</td>
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<td>3.08</td>
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<tr>
<td></td>
<td></td>
<td>48</td>
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<td>3.96</td>
</tr>
<tr>
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</tr>
<tr>
<td></td>
<td></td>
<td>48</td>
<td>3.13</td>
<td>+0.08</td>
<td>+3.27</td>
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<tr>
<td>20509 n/m</td>
<td>( 10^6 )</td>
<td>6</td>
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</tr>
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<td>48</td>
<td>1.42</td>
<td>-0.15</td>
<td>+0.47</td>
<td>+0.24</td>
<td>-6.98</td>
</tr>
</tbody>
</table>

* Col, colistin; Imi, imipenem. A gray background indicates activity (a reduction of >1 log_{10} CFU/ml below the initial inoculum); a green background indicates synergy (a >2-log_{10} decrease in the number of CFU/ml with the combination from that with its most active component); and a red background indicates additivity (a 1.0- to <2-log_{10} decrease in the number of CFU/ml with the combination from that with its most active component).
and imipenem at the commencement of therapy may help to quickly reduce bacterial levels so as to facilitate clearance by the immune system.

Previous time-kill studies have examined colistin in combination with carbapenems against *P. aeruginosa* (2, 9, 38, 39, 46). These studies examined colistin with imipenem, meropenem, or doripenem at a single inoculum (10^6 or 10^7 CFU/ml), though the emergence of colistin resistance was not examined (e.g., by use of PAPs). The present study is the first to investigate the emergence of colistin resistance with colistin combination therapy. In the present investigations, in cases where the combination led to extensive killing at 48 h, meaningful interpretation of the PAPs was not possible (e.g., Fig. 2B, colistin at 4× MIC as monotherapy and in combination with imipenem at 4× MIC). When bacterial numbers at 48 h were comparable, changes in PAPs with combination therapy generally mirrored those observed with equivalent exposure to colistin as monotherapy. However, in both the present study and previously reported studies (2, 9, 38, 39, 46), static concentrations and the instability of carbapenems in aqueous media may have contributed to regrowth and the emergence of colistin resistance at 48 h (20). Thus, it will be important to further assess the utility of these combinations against a range of isolates with various susceptibilities (including heteroresistant strains) in dynamic *in vitro* models and *in vivo*.

Two possible reasons for the enhanced pharmacodynamic effect observed with the combination of colistin and imipenem are subpopulation synergy and mechanistic synergy, as proposed previously (7). Subpopulation synergy involves one drug killing the subpopulation(s) resistant to the other drug, and *vice versa*. Four of the six isolates in the present study were colistin heteroresistant (Table 1), indicating the existence of colistin-resistant subpopulations prior to therapy. In addition, the four imipenem-susceptible isolates were imipenem heteroresistant (while the MIC was 4 mg/liter, subpopulations grew in the presence of 4 mg/liter imipenem [data not shown]).

Another possibility is mechanistic synergy, whereby colistin and imipenem, acting on different cellular pathways, each increase the rate or extent of killing of the other drug. In Gram-negative bacteria, carbapenems must first gain entry into the periplasmic space in order to bind to critical penicillin-binding proteins located on the cytoplasmic membrane (37, 55). A number of resistance mechanisms may operate to limit the concentration of carbapenems in the periplasm, including the presence of carbapenem-hydrolyzing enzymes and the loss of outer membrane proteins (55). Polymyxins cause considerable permeabilization of the outer membrane (57). It is possible that the effect of colistin on membrane permeability results in substantially increased concentrations of imipenem in the periplasm and improved bactericidal activity. Subpopulation and mechanistic synergies are not mutually exclusive; both may operate simultaneously. Further studies, including mechanism-based mathematical modeling, to investigate the mechanism(s)

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Inoculum (CFU/mL)</th>
<th>Time (h)</th>
<th>Log change [log_{10}(CFU)/ml]</th>
<th>Log change [log_{10}(CFU)/ml]</th>
</tr>
</thead>
<tbody>
<tr>
<td>Col resistant, Imi susceptible</td>
<td>32</td>
<td>0.5× MIC</td>
<td>Col 1.0× MIC</td>
<td>Imi 4× MIC</td>
</tr>
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<td>19147 n/m</td>
<td>~10^6</td>
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<td>-0.08</td>
<td>-0.35</td>
</tr>
<tr>
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<td>+2.50</td>
</tr>
<tr>
<td>Col resistant, Imi susceptible</td>
<td>32</td>
<td>0.5× MIC</td>
<td>Col 1.0× MIC</td>
<td>Imi 4× MIC</td>
</tr>
<tr>
<td>19271 n/m</td>
<td>~10^6</td>
<td>6</td>
<td>-1.89</td>
<td>-3.32</td>
</tr>
<tr>
<td></td>
<td>24</td>
<td>+0.49</td>
<td>+2.19</td>
<td>+2.95</td>
</tr>
<tr>
<td></td>
<td>48</td>
<td>+2.87</td>
<td>+1.83</td>
<td>+3.01</td>
</tr>
</tbody>
</table>

**TABLE 3.** Log changes in viable cell counts at 6, 24, and 48 h with various clinically relevant concentrations of colistin and imipenem against one colistin-resistant, imipenem-susceptible isolate and two colistin-susceptible, imipenem-resistant isolates of *P. aeruginosa*.

Col, colistin; Imi, imipenem. A gray background indicates activity (a reduction of ≥1 log_{10} CFU/ml below the initial inoculum); a green background indicates synergy (a ≥2 log_{10} decrease in the number of CFU/ml with the combination from that with its most active component); and a red background indicates additivity (a 1.0- to <2-log_{10} decrease in the number of CFU/ml with the combination from that with its most active component). For colistin-resistant isolate 19147 n/m, synergy or additivity was compared with imipenem monotherapy only.
underpinning the enhanced pharmacodynamic activity observed are ongoing.

In the battle against rapidly emerging bacterial resistance in Gram-negative “superbugs,” rational approaches to the use of combinations of existing antibiotics may be very beneficial. To the best of our knowledge, this is the first systematic study on the PD of colistin in combination with imipenem against P. aeruginosa, including MDR and colistin-heteroresistant strains, at both low and high inocula. Clinically relevant concentrations of colistin in combination with imipenem substantially increased bacterial killing against MDR P. aeruginosa isolates at both inocula when isolates were resistant to either antibiotic. Further investigations in in vitro pharmacodynamic systems, animal infection models, and clinical studies are warranted to optimize colistin-imipenem combinations targeting both colistin-susceptible and colistin-resistant subpopulations.

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